Bioinformatics 3 V 4 – Weak Indicators and Communities

Mon, Oct 28, 2013

#### Noisy Data — Clear Statements?

For **yeast**: ~ 6000 proteins  $\rightarrow$  ~18 million potential interactions rough estimates:  $\leq$  100000 interactions occur

- $\rightarrow$  1 true positive for 200 potential candidates = **0.5%** 
  - $\rightarrow$  decisive experiment must have accuracy << 0.5% false positives

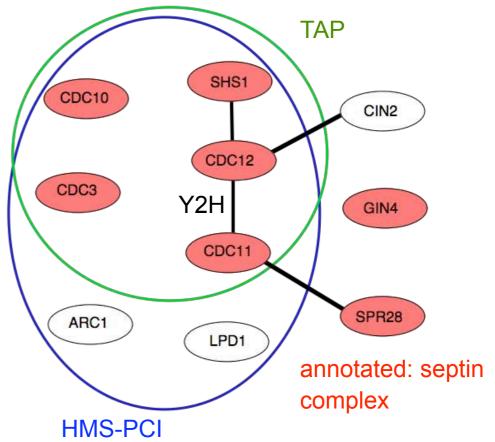
#### **Different experiments** detect different interactions

For yeast: 80000 interactions known, only 2400 found in > 1 experiment

Y2H:  $\rightarrow$  many false positives (up to 50% errors)

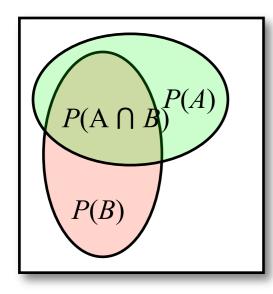
Co-expression:  $\rightarrow$  gives indications at best

Combine weak indicators = ???



## **Conditional Probabilities**

Joint probability for "*A* and *B*":



 $P(A \cap B) = P(A|B) P(B) = P(B|A) P(A)$ 

Solve for conditional probability for "A when B is true"  $\rightarrow$  Bayes' Theorem:

$$P(A|B) = \frac{P(B|A) P(A)}{P(B)} = \frac{P(B|A)}{P(B)} P(A)$$

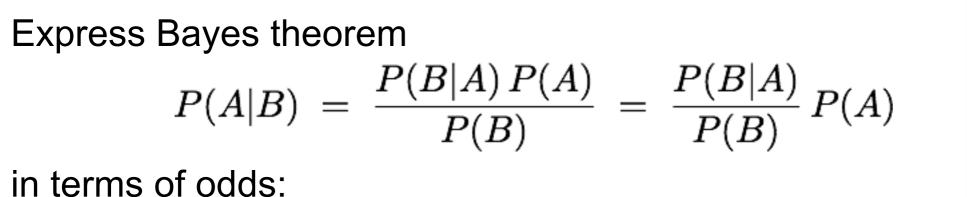
P(A) = prior probability (marginal prob.) for "A"  $\rightarrow$  no prior knowledge about A

P(B) = prior probability for "B"  $\rightarrow$  normalizing constant

- P(B | A) = conditional probability for "B given A"
- P(A | B) = posterior probability for "A given B"

 $\rightarrow$  Use information about *B* to improve knowledge about *A* 

## What are the Odds?



 $P(A \cap B)$  P(A) P(B)

- Also Consider case "A does not apply":  $P(\bar{A}|B) = \frac{P(B|A)}{P(B)}P(\bar{A})$
- odds for A when we know about B
  (we will interpret B as information or features):

$$O(A|B) = \frac{P(A|B)}{P(\overline{A}|B)} = \frac{P(B|A)}{P(B|\overline{A})} \frac{P(A)}{P(\overline{A})} = \Lambda(A|B) O(A)$$
posterior odds for A likelihood ratio prior odds for A

 $\Lambda(A \mid B) \rightarrow$  by how much does our knowledge about A improve?

## 2 types of Bayesian Networks

Encode conditional dependencies between evidences



"A depends on B"
 with the conditional probability P(A | B)

Evidence nodes can have a variety of types: numbers, categories, ...

#### (1) Naive Bayesian network

 $\rightarrow$  independent odds  $O(A|B,C) = \Lambda(A|B) \Lambda(A|C) O(A)$ 

(2) Fully connected Bayesian network		В	!B	
$\rightarrow$ table of joint odds	С	0.3	0.16	$\Leftrightarrow \Lambda(A B,C)$
	!C	0.4	0.14	,,(,2,0)

### **Bayesian Analysis of Complexes**

#### A Bayesian Networks Approach for Predicting Protein-Protein Interactions from Genomic Data

Ronald Jansen,<sup>1</sup>\* Haiyuan Yu,<sup>1</sup> Dov Greenbaum,<sup>1</sup> Yuval Kluger,<sup>1</sup> Nevan J. Krogan,<sup>4</sup> Sambath Chung,<sup>1,2</sup> Andrew Emili,<sup>4</sup> Michael Snyder,<sup>2</sup> Jack F. Greenblatt,<sup>4</sup> Mark Gerstein<sup>1,3</sup>†

We have developed an approach using Bayesian networks to predict proteinprotein interactions genome-wide in yeast. Our method naturally weights and combines into reliable predictions genomic features only weakly associated with interaction (e.g., messenger RNA coexpression, coessentiality, and colocalization). In addition to de novo predictions, it can integrate often noisy, experimental interaction data sets. We observe that at given levels of sensitivity, our predictions are more accurate than the existing high-throughput experimental data sets. We validate our predictions with TAP (tandem affinity purification) tagging experiments. Our analysis, which gives a comprehensive view of yeast interactions, is available at genecensus.org/intint.

Science 302 (2003) 449

## Improving the Odds

Is a given protein pair AB a complex (from all that we know)?

 $O_{post}(\text{Complex}|f_1, f_2, \dots) = \Lambda(\text{Complex}|f_1, f_2, \dots) O_{prior}(\text{Complex})$ 

likelihood ratio: improvement of the odds when we know about features f1, f2, 1 ... idea: determine from known complexes and use for prediction of new complexes

```
prior odds for a
random pair AB to be
a complex
f
estimate (somehow)
```

Features used by Jansen et al (2003):

- 4 experimental data sets of complexes
- mRNA co-expression profiles
- biological functions annotated to the proteins (GO, MIPS)
- essentiality for the cell

## **Gold Standard Sets**

To determine  $\Lambda(\text{Complex}|f_1, f_2, \dots) = \frac{P(f_1, f_2, \dots | \text{Complex})}{P(f_1, f_2, \dots | \text{no Complex})}$ 

 $\rightarrow$  use two data sets with **known** features  $f_1, f_2, \dots$  for **training** 

Requirements for training data:

- i) independent of the data serving as evidence
- ii) large enough for good statistics
- iii) free of systematic bias

#### Gold Standard Positive Set (GP):

8250 complexes from the hand-curated MIPS catalog of protein complexes (MIPS stands for Munich Information Center for Protein Sequences)

#### Gold Standard Negative Set (GN):

2708746 (non-)complexes formed by proteins from different cellular compartments (assuming that such protein pairs likely do not interact)

$$Prior Odds$$

$$O_{prior}(Complex) = \frac{P(Complex)}{P(no Complex)} = \frac{P(Complex)}{1 - P(Complex)}$$

Jansen et al:

- estimated  $\geq$  30000 existing complexes in yeast
- 18 Mio. possible complexes

 $\rightarrow$  *P*(Complex)  $\approx$  1/600

 $\rightarrow$  O<sub>prior</sub> = 1/600

- $\rightarrow$  The odds are 600 : 1 against picking a complex at random
- $\rightarrow$  expect 50% good hits (TP > FP) with  $\Lambda \approx 600$

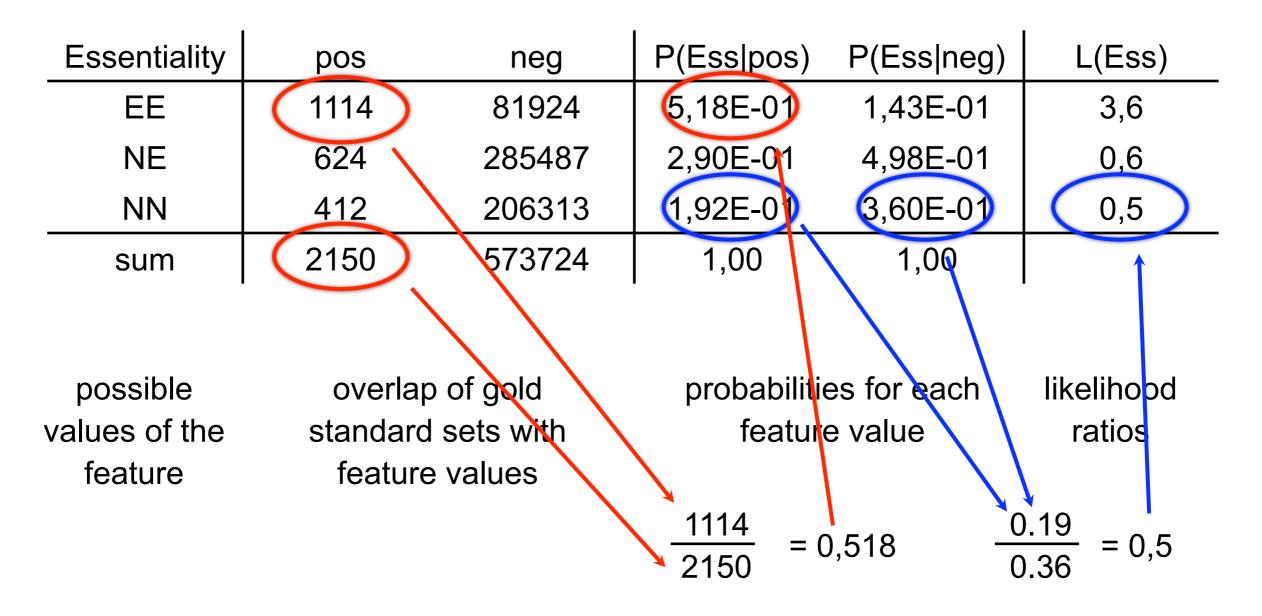
Note: Oprior is mostly an educated guess

## Essentiality

Test whether both proteins are essential (E) for the cell or not (N)  $\rightarrow$  for protein complexes, EE or NN should occur more often

pos/neg: # of gold standard positives/ negatives with essentiality information

$$L(\text{Ess}) = \frac{P(\text{Ess} | \text{pos})}{P(\text{Ess} | \text{neg})}$$



# mRNA Co-Expression

Publicly available expression data from

- the Rosetta compendium
- the yeast cell cycle

Correlation between the data sets  $\rightarrow$  use principal component

			Gold standa	ard overlap						
	Expression correlation	# protein pairs	pos	neg	sum(pos)	sum( <i>neg</i> )	<pre>sum(pos )/ P(exp pos sum(neg )</pre>		P(exp neg)	L
	0.9	678	16	45	16	45	0.36	2.10E-03	1.68E-05	124.9
	0.8	4,827	137	563	153	608	0.25	1.80E-02	2.10E-04	85.5
	0.7	17,626	530	2,117	683	2,725	0.25	6.96E-02	7.91E-04	88.0
	0.6	42,815	1,073	5,597	1,756	8,322	0.21	1.41E-01	2.09E-03	67.4
	0.5	96,650	1,089	14,459	2,845	22,781	0.12	1.43E-01	5.40E-03	26.5
	0.4	225,712	993	35,350	3,838	58,131	0.07	1.30E-01	1.32E-02	9.9
	0.3	529,268	1,028	83,483	4,866	141,614	0.03	1.35E-01	3.12E-02	4.3
	0.2	1,200,331	870	183,356	5,736	324,970	0.02	1.14E-01	6.85E-02	1.7
es	0.1	2,575,103	739	368,469	6,475	693,439	0.01	9.71E-02	1.38E-01	0.7
Values	0	9,363,627	894	1,244,477	7,369	1,937,916	0.00	1.17E-01	4.65E-01	0.3
Š	-0.1	2,753,735	164	408,562	7,533	2,346,478	0.00	2.15E-02	1.53E-01	0.1
	-0.2	1,241,907	63	203,663	7,596	2,550,141	0.00	8.27E-03	7.61E-02	0.1
	-0.3	484,524	13	84,957	7,609	2,635,098	0.00	1.71E-03	3.18E-02	0.1
	-0.4	160,234	3	28,870	7,612	2,663,968	0.00	3.94E-04	1.08E-02	0.0
	-0.5	48,852	2	8,091	7,614	2,672,059	0.00	2.63E-04	3.02E-03	0.1
	-0.6	17,423	-	2,134	7,614	2,674,193	0.00	0.00E+00	7.98E-04	0.0
	-0.7	7,602	-	807	7,614	2,675,000	0.00	0.00E+00	3.02E-04	0.0
	-0.8	2,147	-	261	7,614	2,675,261	0.00	0.00E+00	9.76E-05	0.0
	-0.9	67	-	12	7,614	2,675,273	0.00	0.00E+00	4.49E-06	0.0
	Sum	18,773,128	7,614	2,675,273	-	-	-	1.00E+00	1.00E+00	1.0

## **Biological Function**

Use MIPS function catalog and Gene Ontology function annotations

- determine functional class shared by the two proteins; small values (1-9)
   Indicate highest MIPS function or GO BP similarity
- count how many of the 18 Mio potential pairs share this classification

			Gold stand	ard overlap						
	MIPS function similarity	# protein pairs	pos	neg	sum( <i>pos</i> )	sum(neg)	sum(pos )/ sum(neg )	P(MIPS pos)	P(MIPS neg)	L
	1 9	6,584	171	1,094	171	1,094	0.16	2.12E-02	8.33E-04	25.5
es	10 99	25,823	584	4,229	755	5,323	0.14	7.25E-02	3.22E-03	22.5
Value	100 1000	88,548	688	13,011	1,443	18,334	0.08	8.55E-02	9.91E-03	8.6
Š	1000 10000	255,096	6,146	47,126	7,589	65,460	0.12	7.63E-01	3.59E-02	21.3
	10000 Inf	5,785,754	462	1,248,119	8,051	1,313,579	0.01	5.74E-02	9.50E-01	0.1
	Sum	6,161,805	8,051	1,313,579	-	-	-	1.00E+00	1.00E+00	1.0

			Gold stand	ard overlap						
GO	biological process similarity	# protein pairs	pos	neg	sum(pos)	sum( <i>neg</i> )	sum(pos )/ sum(neg )	P(GO pos)	P(GO neg)	L
	1 9	4,789	88	819	88	819	0.11	1.17E-02	1.27E-03	9.2
es	10 99	20,467	555	3,315	643	4,134	0.16	7.38E-02	5.14E-03	14.4
alue	100 1000	58,738	523	10,232	1,166	14,366	0.08	6.95E-02	1.59E-02	4.4
2	1000 10000	152,850	1,003	28,225	2,169	42,591	0.05	1.33E-01	4.38E-02	3.0
	10000 Inf	2,909,442	5,351	602,434	7,520	645,025	0.01	7.12E-01	9.34E-01	0.8
	Sum	3,146,286	7,520	645,025	-	-	-	1.00E+00	1.00E+00	1.0

### **Experimental Data Sets**

In vivo pull-down: Gavin et al, *Nature* **415** (2002) 141 31304 pairs Ho et al, *Nature* **415** (2002) 180 25333 pairs

HT-Y2H:Uetz et al, Nature 403 (2000) 623981 pairsIto et al, PNAS 98 (2001) 45694393 pairs

4 experiments on overlapping PP pairs

 $\rightarrow$  2<sup>4</sup> = 16 categories — fully connected Bayes network

Caulin		11-4-	14.0	# nrotoin		Gold-standard overlap						
Gavin (g)	Ho (h)	(u)	(i)	# protein pairs	pos	neg	sum(pos)	sum(neg)	sum(pos)/ sum(neg)	P(g,h,u,i   pos)	P(g,h,u,i   neg)	L
1	1	1	0	16	6	0	6	0	-	7.27E-04	0.00E+00	-
1	0	0	1	53	26	2	32	2	16.0	3.15E-03	7.38E-07	4268.3
1	1	1	1	11	9	1	41	3	13.7	1.09E-03	3.69E-07	2955.0
1	0	1	1	22	6	1	47	4	11.8	7.27E-04	3.69E-07	1970.0
1	1	0	1	27	16	3	63	7	9.0	1.94E-03	1.11E-06	1751.1
1	0	1	0	34	12	5	75	12	6.3	1.45E-03	1.85E-06	788.0
1	1	0	0	1920	337	209	412	221	1.9	4.08E-02	7.72E-05	529.4
0	1	1	0	29	5	5	418	227	1.8	6.06E-04	1.85E-06	328.3
0	1	1	1	16	1	1	413	222	1.9	1.21E-04	3.69E-07	328.3
0	1	0	1	39	3	4	421	231	1.8	3.64E-04	1.48E-06	246.2
0	0	1	1	123	6	23	427	254	1.7	7.27E-04	8.49E-06	85.7
1	0	0	0	29221	1331	6224	1758	6478	0.3	1.61E-01	2.30E-03	70.2
0	0	1	0	730	5	112	1763	6590	0.3	6.06E-04	4.13E-05	14.7
0	0	0	1	4102	11	644	1774	7234	0.2	1.33E-03	2.38E-04	5.6
0	1	0	0	23275	87	5563	1861	12797	0.1	1.05E-02	2.05E-03	5.1
0	0	0	0	2702284	6389	2695949	8250	2708746	0.0	7.74E-01	9.95E-01	0.8

### **Statistical Uncertainties**

Carrie	11-	11-4-	14-	#		Gold				
Gavin (g)	но (h)	(u)	(i)	# protein pairs	pos	neg	P(g,h,u,i   pos)	P(g,h,u,i   neg)	L	
1	1	1	0	16	6	0	7.27E-04	0.00E+00	-	
1	0	0	1	53	26	2	3.15E-03	7.38E-07	4268.3	
1	1	1	1	11	9	1	1.09E-03	3.69E-07	2955.0	
1	0	1	1	22	6	1	7.27E-04	3.69E-07	1970.0	
1	1	0	1	27	16	3	1.94E-03	1.11E-06	1751.1	
1	0	1	0	34	12	5	1.45E-03	1.85E-06	788.0	

1) L(1111) < L(1001)

statistical uncertainty:  $\Delta N = \sqrt{N+1}$ 

Overlap with all experiments is smaller  $\rightarrow$  larger uncertainty

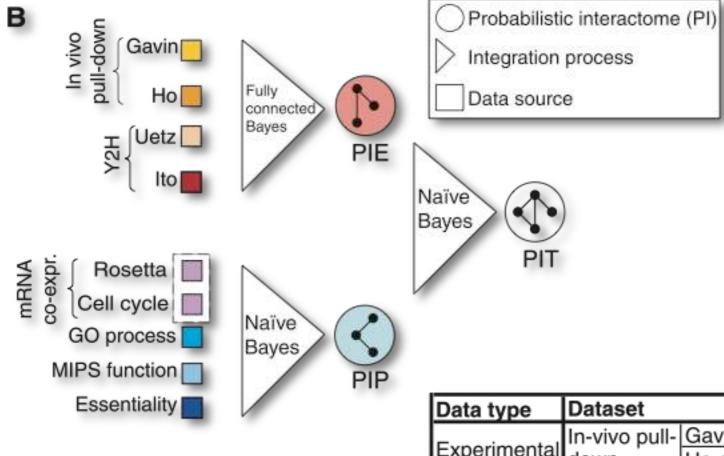
2) *L*(1110) = NaN?

Use conservative lower bound  $\rightarrow$  assume 1 overlap with GN  $\rightarrow L(1110) \ge 1970$ 

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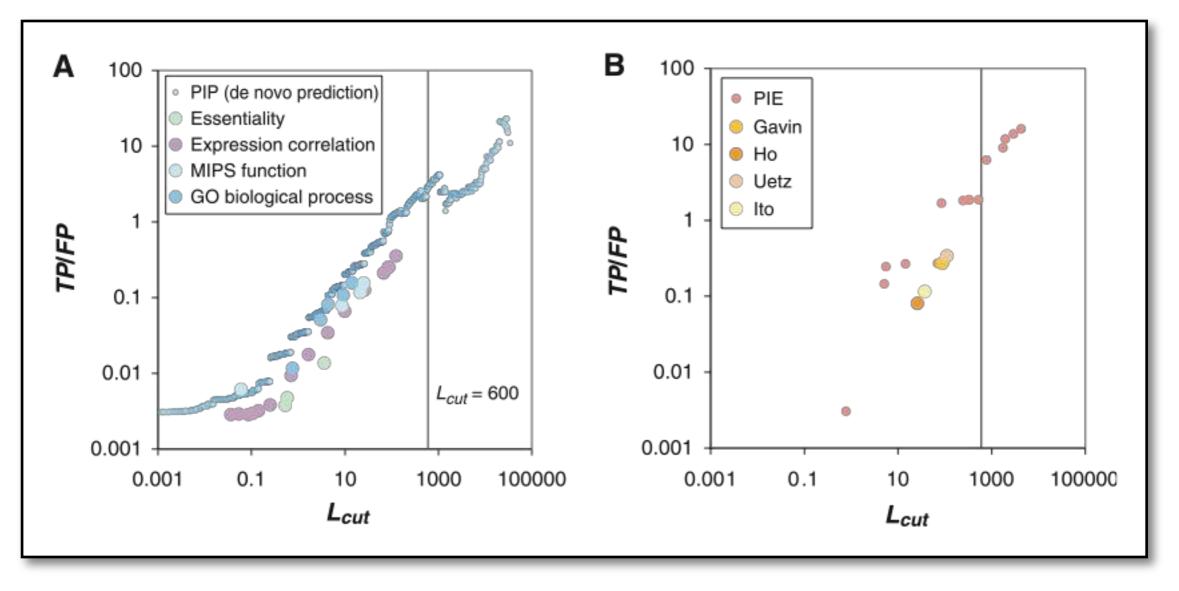
Jansen et al, Science 302 (2003) 449

### Overview



Data type	Dataset			# protein pairs	Used for	
Experimental	In-vivo pull-	Gavin et al.		31,304	Integration of	
Experimental interaction	down	Ho et al.			experimental	
data	Yeast two-	Uetz et al.			interaction	
uala	hybrid	Ito et al.		4,393	data (PIE)	
	mRNA	Rosetta compendium		19,334,806		
Other	Expression	Cell cycle		17,467,005	De novo	
genomic	Biological	GO biological process		3,146,286	prediction	
features	function	MIPS function		6,161,805	(PIP)	
	Essentiality			8,130,528		
Gold	Positives	Proteins in the same MIPS complex		8,250	Training &	
standards	Negatives	Proteins separated by localization		2,708,746	testing	

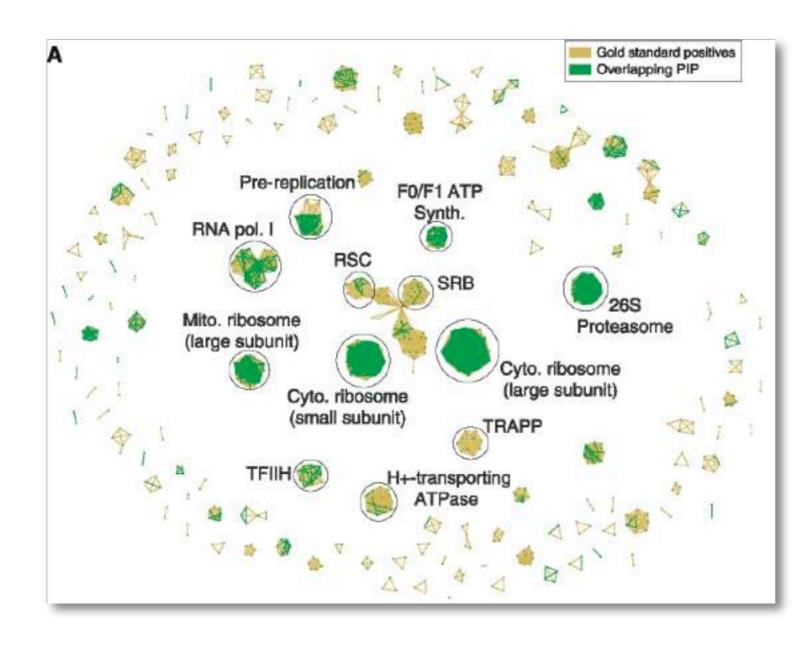
# Performance of complex prediction



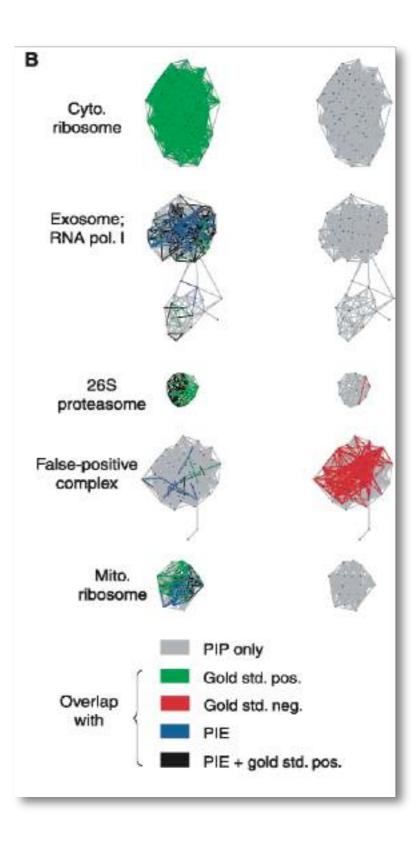
Re-classify Gold standard complexes:
Ratio of true positives to false positives
→ None of the evidences alone was enough

$$\frac{TP}{FP}(L_{cut}) = \frac{\sum_{L>L_{cut}} \operatorname{pos}(L)}{\sum_{L>L_{cut}} \operatorname{neg}(L)}$$

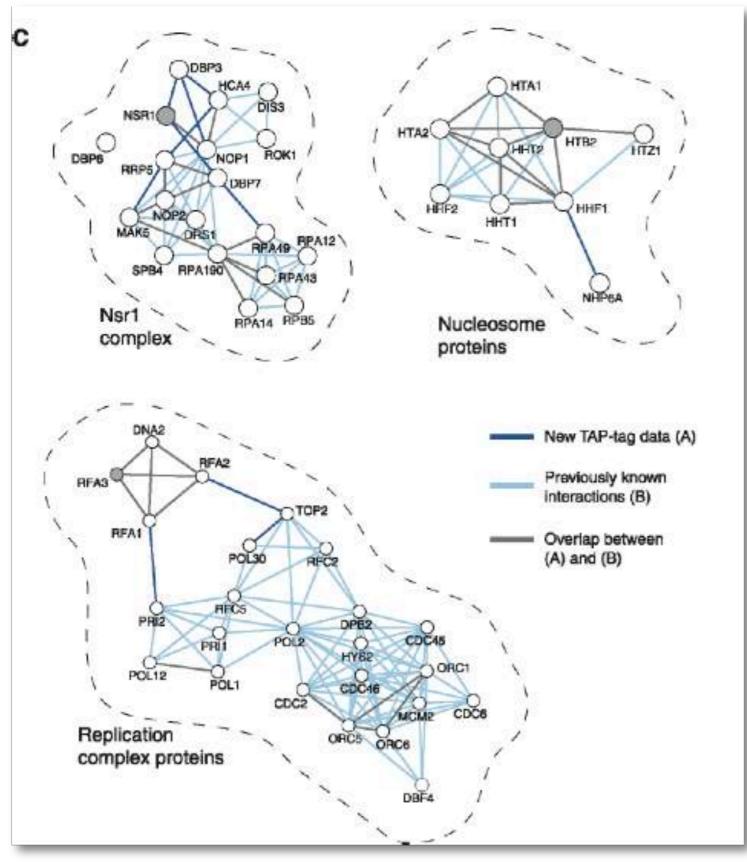
## Coverage



Predicted set covers 27% of the GP



## **Verification of Predicted Complexes**



Compare predicted complexes with available experimental evidence and directed new TAPtag experiments

→ use directed
 experiments to verify
 new predictions
 (more efficient)

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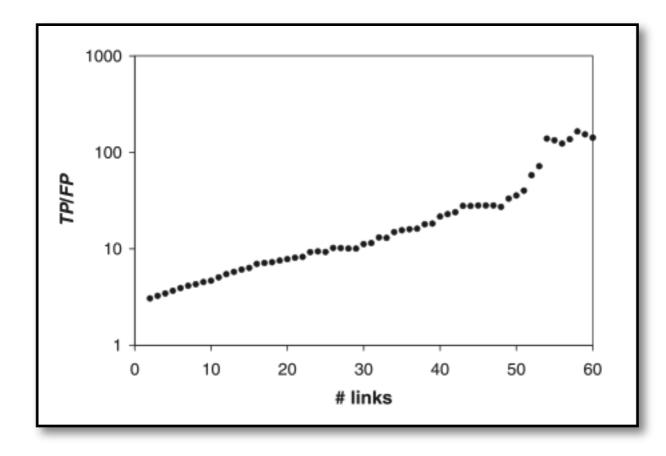
Jansen et al, Science 302 (2003) 449

### **Consider Connectivity**

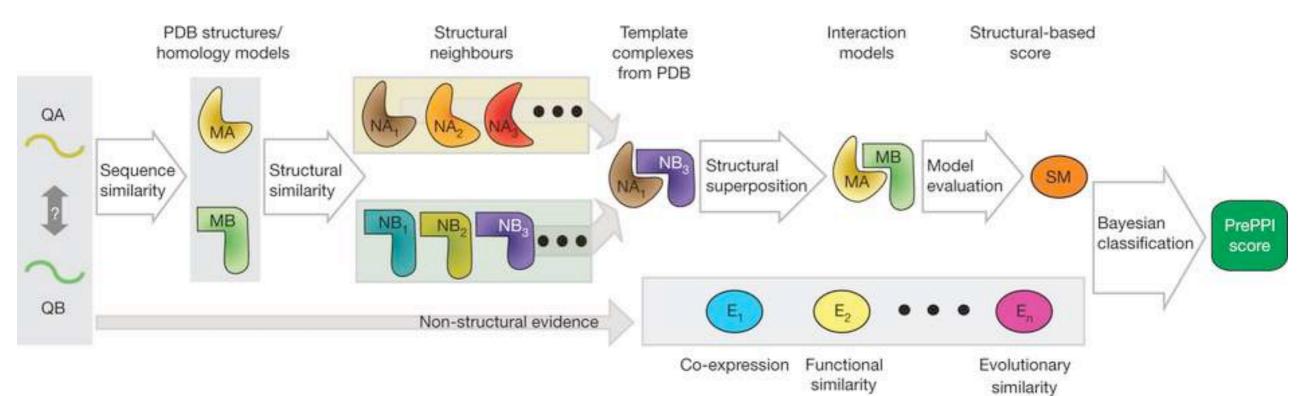
Only take proteins e.g. with  $\geq$  20 links

 $\rightarrow$  This preserves links inside the complex,

filter false-positive links to heterogeneous groups outside the complex



# Follow-up work: PrePPI (2012)



Given a pair of query proteins that potentially interact (QA, QB), representative structures for the individual subunits (MA, MB) are taken from the PDB, where available, or from homology model databases.

For each subunit we find both close and remote structural neighbours. A 'template' for the interaction exists whenever a PDB or PQS structure contains a pair of interacting chains (for example,  $NA_1-NB_3$ ) that are structural neighbours of MA and MB, respectively. A model is constructed by superposing the individual subunits, MA and MB, on their corresponding structural neighbours,  $NA_1$  and  $NB_3$ .

We assign 5 empirical-structure-based scores to each interaction model and then calculate a likelihood for each model to represent a true interaction by combining these scores using a Bayesian network trained on the HC and the N interaction reference sets.

We finally combine the structure-derived score (SM) with non-structural evidence associated with the query proteins (for example, co-expression, functional similarity) using a naive Bayesian classifier.

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Zhang et al, Nature (2012) 490, 556–560

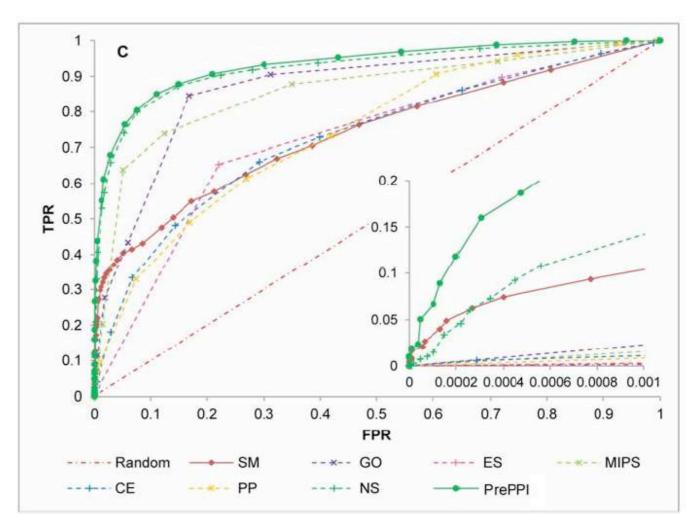
### **Results of PrePPI**

Receiver-operator characteristics (ROC) for predicted yeast complexes.

Examined features:

- structural modeling (SM),
- GO similarity,
- protein essentiality (ES) relationship,
- MIPS similarity,
- co-expression (CE),
- phylogenetic profile (PP) similarity.

Also listed are 2 combinations: - NS for the integration of all non-structure clues, i.e. GO, ES, MIPS, CE, and PP, and - PrePPI for all structural and non-structure clues).



This gave 30.000 high-confidence PP interactions for yeast and 300.000 for human.

# Summary: Bayesian Analysis

Combination of weak features yields powerful predictions

- boosts odds via Bayes' theorem
- Gold standard sets for training the likelihood ratios

Bayes vs. other **machine learning** techniques: (voting, unions, SVM, neuronal networks, decision trees, ...)

- $\rightarrow$  arbitrary types of data can be combined
- $\rightarrow$  weight data according to their **reliability**
- $\rightarrow$  include conditional relations between evidences
- $\rightarrow$  easily accommodates missing data (e.g., zero overlap with GN)
- → **transparent** procedure
- $\rightarrow$  predictions easy to **interpret**

# **Connected Regions**

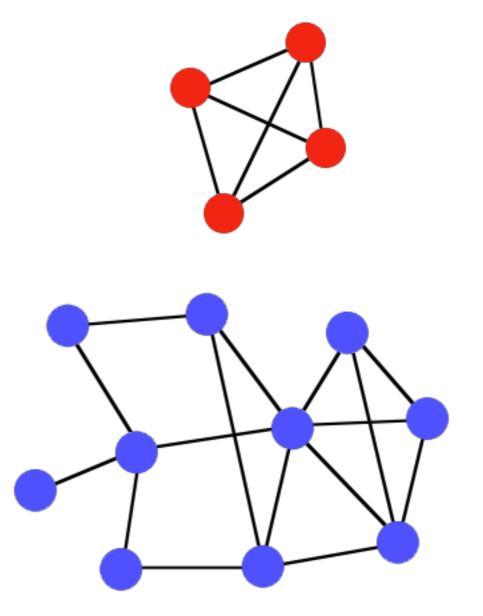
Observation: more interactions inside a complex than to the outside

 $\rightarrow$  how can one identify highly connected regions in a network?

1) Fully connected region: **Clique** clique :=  $G' = (V', E' = V^{(2)})$ 

#### Problems with cliques:

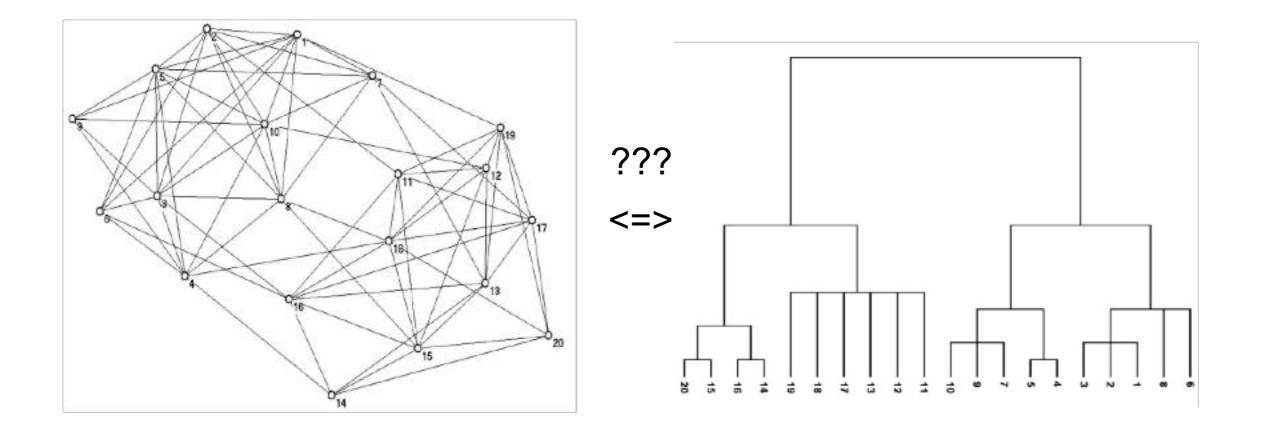
- finding cliques is NP-hard (but "works" O(N<sup>2</sup>) for the sparsely connected biological networks)
- biological protein complexes are not always fully connected



## Communities

Community := subset of vertices, for which the **internal** connectivity is **denser** than to the outside

Aim: map network onto tree that reflects the community structure



Radicchi et al, *PNAS* **101** (2004) 2658:

## **Hierarchical Clustering**

- 1) Assign a weight *W<sub>ij</sub>* to each pair of vertices *i*, *j* that measures how "closely related" these two vertices are.
- 2) Iteratively add edges between pairs of nodes with decreasing  $W_{ij}$

#### **Measures** for *W*<sub>ij</sub>:

1) Number of **vertex-independent paths** between vertices *i* and *j* (vertex-independent paths between *i* and *j*: no shared vertex except *i* and *j*)

Menger (1927): the number of vertex-independent paths equals the number of vertices that have to be removed to cut all paths between *i* and *j*  $\rightarrow$  measure for network robustness

- 2) Number of **edge-independent paths** between *i* and *j*
- 3) **Total number of paths** *L* between *i* and *j* but L = 0 or  $\infty \rightarrow$  weight paths with their length  $\alpha^{L}$  with  $\alpha < 1$

Problem: vertices with a single link are separated from the communities

#### Vertex Betweenness

Freeman (1927): count on how many shortest paths a vertex is visited

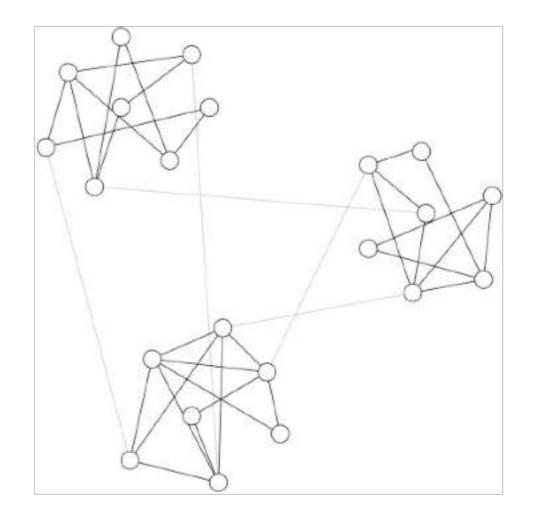
For a graph G = (V, E) with |V| = n

**Betweenness** for vertex v:

$$C_B(\nu) = \frac{\sum_{s \neq \nu \neq t \in V} \sigma_{st}(\nu)}{(n-1)(n-2)}$$

#### Alternative: edge betweenness

→ to how many shortest paths does this edge belong



#### Girvan-Newman Algorithm

Girvan, Newman, PNAS 99 (2002) 7821:

For a graph G = (V, E) with |V| = n, |E| = m

- 1) Calculate **betweenness** for all *m* edges (takes O(*mn*) time)
- 2) **Remove** edge with highest betweenness
- 3) **Recalculate** betweenness for all affected nodes
- 4) **Repeat** from 2) until no more edge is left (at most *n* iterations)
- 5) Build up tree from V by reinserting vertices in reverse order

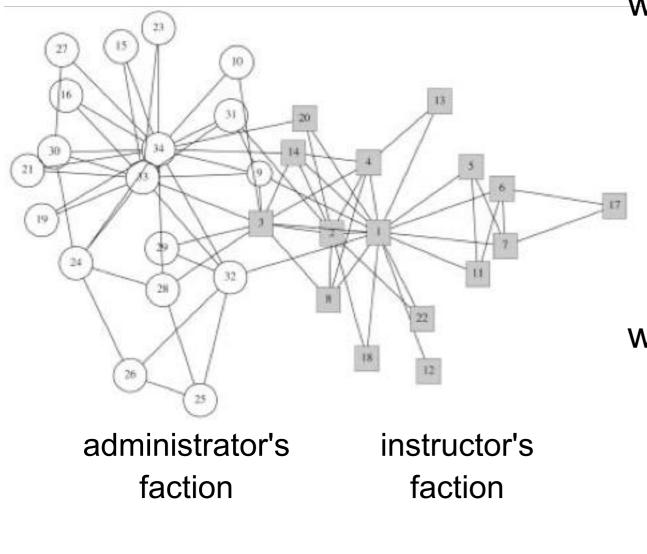
**Works** well, but **slow**:  $O(mn^2) \approx O(n^3)$  for scale-free networks (|E| = 2 |V|)

Reason for complexity: compute shortest paths  $(n^2)$  for *m* edges

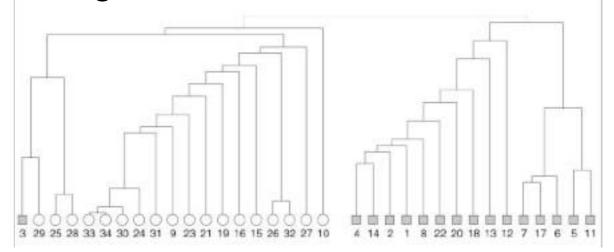
 $\rightarrow$  recalculating a **global** property is expensive for larger networks

## Zachary's Karate Club

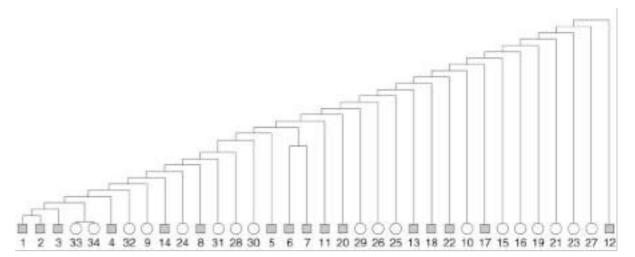
- observed friendship relations of 34 members over two years
- correlate fractions at break-up with calculated communities



with edge betweenness:

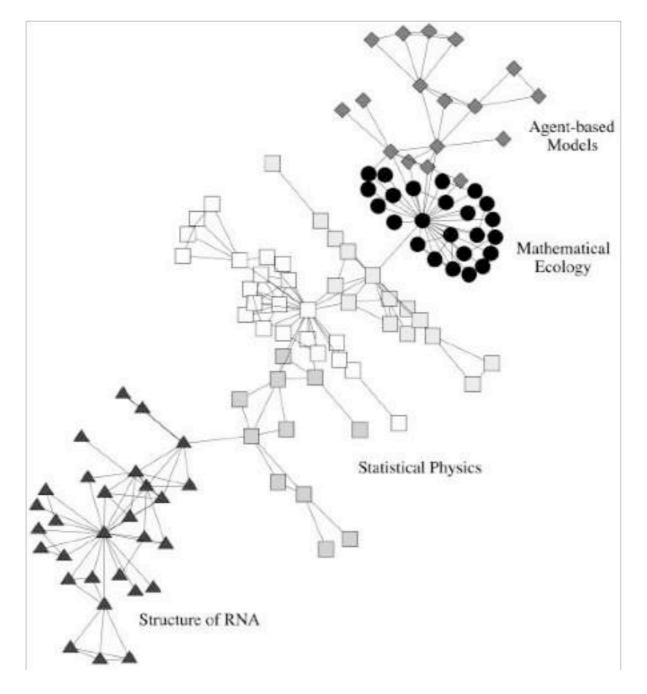


with number of edge-independent paths:



Girvan, Newman, PNAS 99 (2002) 7821

### **Collaboration Network**



The largest component of the Santa Fe Institute collaboration network, with the primary divisions detected by the GN algorithm indicated by different vertex shapes.

Edge: two authors have coauthored a joint paper.

Girvan, Newman, PNAS 99 (2002) 7821

## **Determining Communities Faster**

#### Radicchi et al, PNAS 101 (2004) 2658:

Determine edge weights via edge-clustering coefficient

- $\rightarrow$  local measure
  - $\rightarrow$  much faster, esp. for large networks

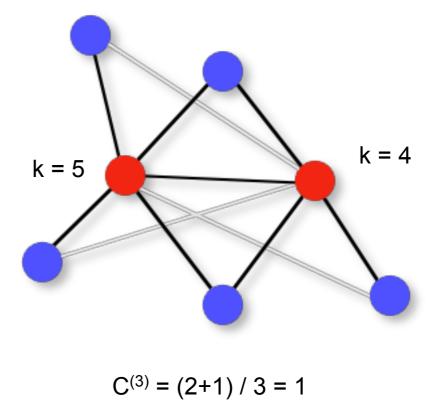
Modified edge-clustering coefficient:

 $\rightarrow$  fraction of potential triangles with edge between *i* and *j* 

$$C_{i,j}^{(3)} = \frac{z_{i,j}^{(3)} + 1}{\min[(k_i - 1), (k_j - 1)]}$$

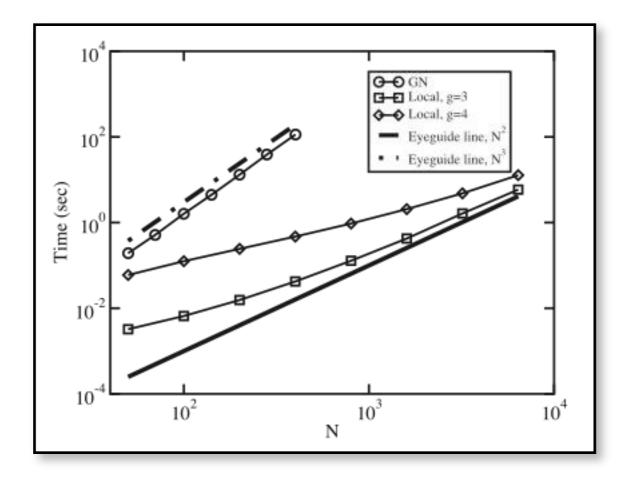
Here,  $z_{i,j}^{(3)}$  is the number of triangles,  $k_i$  and  $k_j$  are the degrees of nodes *i* and *j*.

Note: "+ 1" to remove degeneracy for  $z_{i,j}^{(3)} = 0$ 



#### Performance

Instead of triangles: **cycles** of higher order  $g \rightarrow$  continuous transition to a global measure



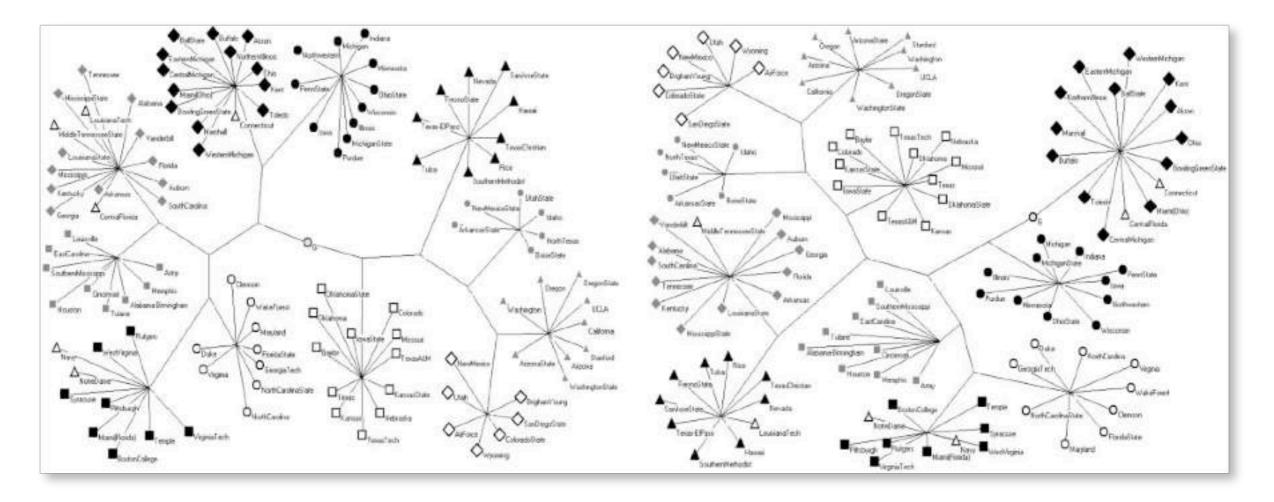
Radicchi et al-algorithm:  $O(N^2)$  for large networks

Radicchi et al, *PNAS* **101** (2004) 2658:

 $C_{i,j}^{(g)}$ 

## **Comparison of algorithms**

Data set: football teams from US colleges; different symbols = different conferences, teams played ca. 7 intraconference games and 4 inter-conference games in 2000 season.



Girven-Newman algorithm

Radicchi with g = 4

 $\rightarrow$  very similar communities

## **Strong Communities**

"Community := subgraph with more interactions inside than to the outside"

A subgraph *V* is a **community** in a...

...strong sense when:

 $k_i^{in}(V) > k_i^{out}(V) \quad \forall i \in V$ 

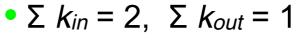
 $\rightarrow$  Check every node individually

...weak sense when:

$$\sum_{i \in V} k_i^{in}(V) > \sum_{i \in V} k_i^{out}(V)$$

 $\rightarrow$  allow for borderline nodes

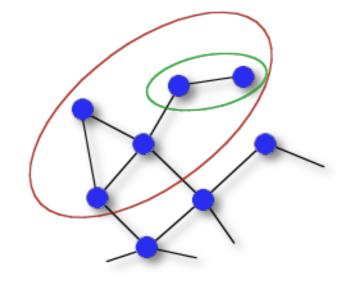
Radicchi et al, *PNAS* **101** (2004) 2658



 $\{k_{in}, k_{out}\} = \{\mathbf{1}, \mathbf{1}\}, \{1, 0\}$ 

 $\rightarrow$  community in a weak sense

Σ k<sub>in</sub> = 10, Σ k<sub>out</sub> = 2 {k<sub>in</sub>, k<sub>out</sub>} = {2,1}, {2, 0}, {3, 1}, {2,0}, {1,0} → community in a strong and weak sense



## Summary

What you learned **today**:

- how to combine a set of noisy evidences into a powerful prediction tool
   → Bayes analysis
- how to find **communities** in a network efficiently
- $\rightarrow$  betweenness, edge-cluster-coefficient

Next lecture: Mon, Nov 4, 2013

- Modular decomposition
- Robustness

#### Short Test #1: Fri, Nov. 8